

CloverDirect™

tRNA Reagents for Site-Directed Protein Functionalization



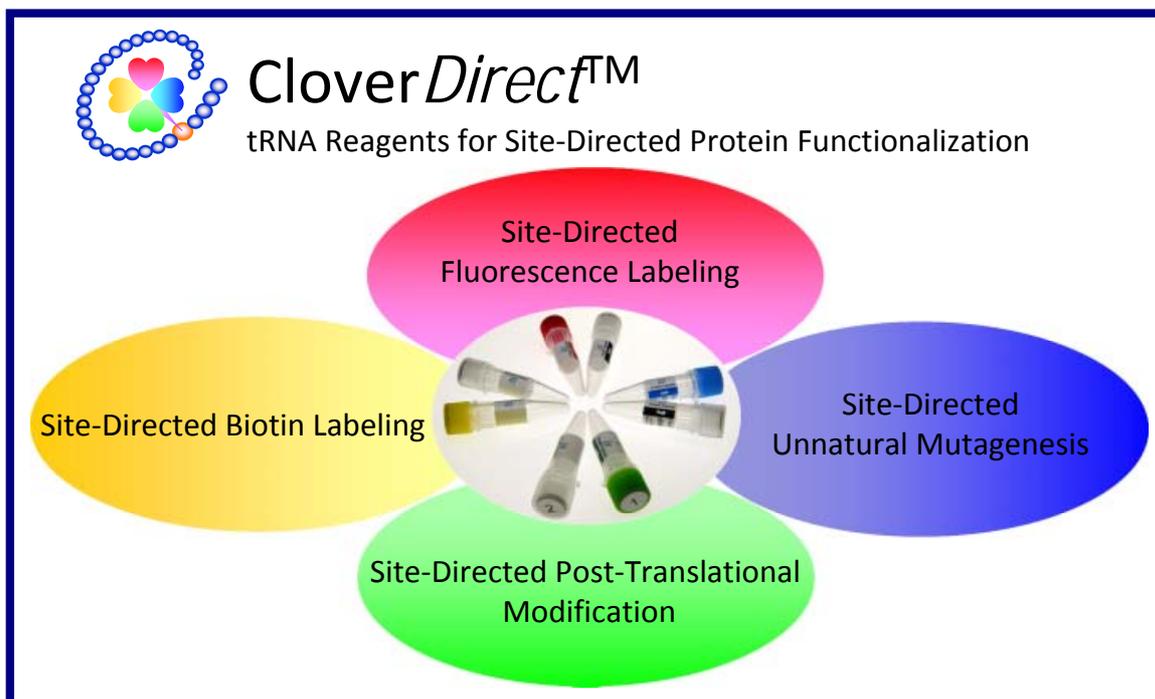


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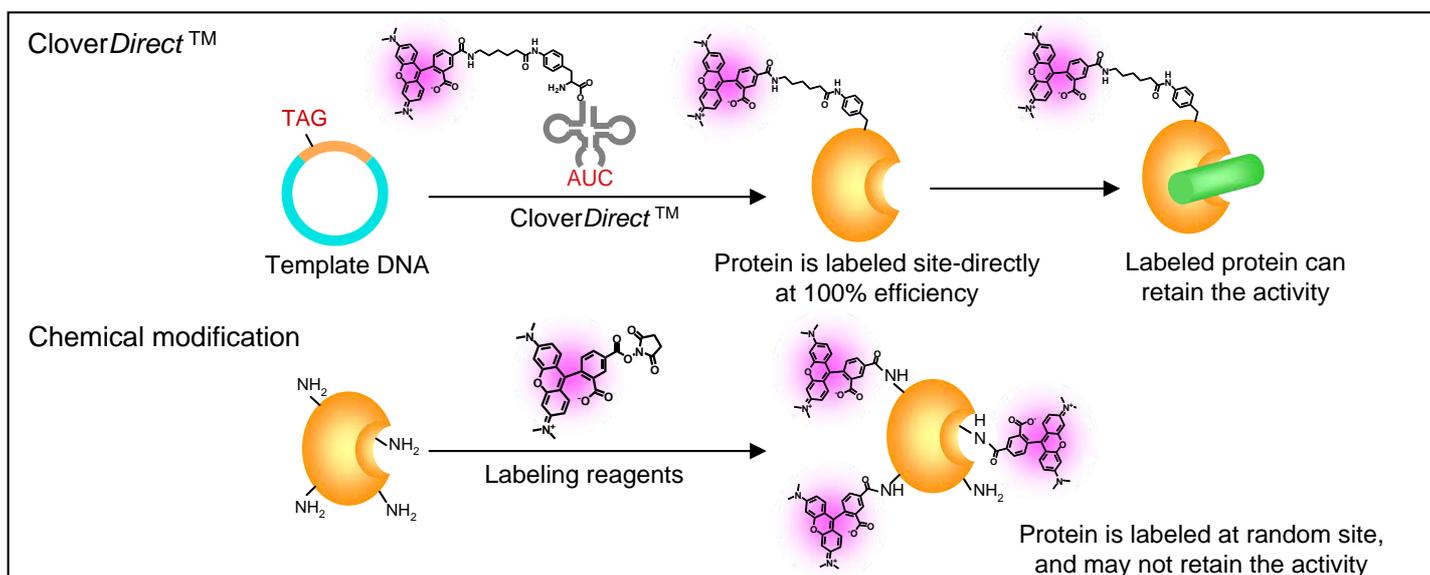
[About CloverDirect™]

CloverDirect™ tRNA Reagents for Site-Directed Protein Functionalization allow the incorporation of unnatural amino acids at defined positions of proteins using *in vitro* translation. Unnatural amino acids containing fluorescent groups, biotin, PEG, photo-crosslink, etc are available. Proteins with unnatural amino acids will be obtained within a few hours just by adding CloverDirect™ reagents and DNA template having an amber stop codon (UAG) or a four-base codon (CGGG) to an *in vitro* translation system. CloverDirect™ covers the following four applications. In addition, we provide custom services for the expression of proteins with unnatural amino acids (see page 11 for details).



Site-Directed Fluorescence Labeling

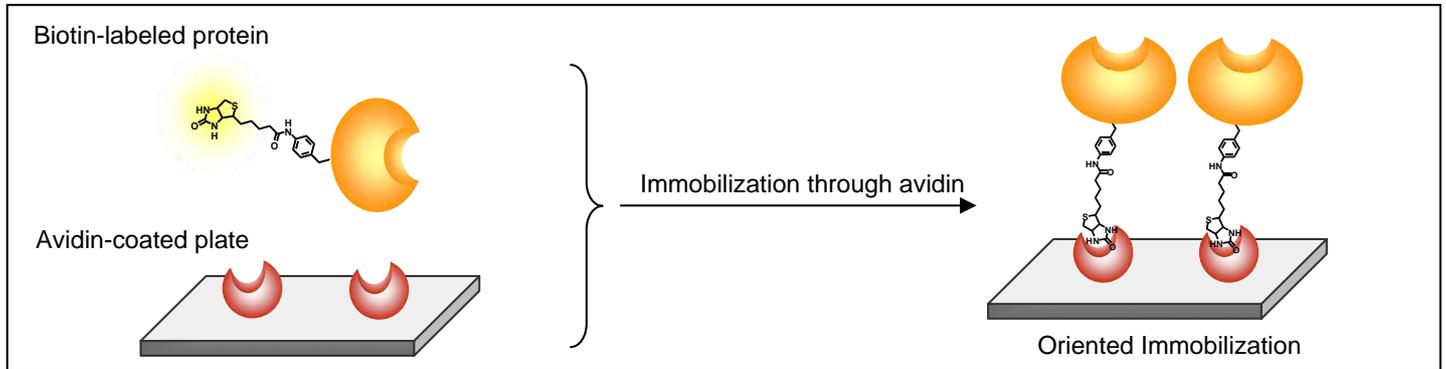
It is not easy to incorporate fluorescent groups into proteins in a site-direct and quantitative fashion by chemical modification. CloverDirect™ tRNA Reagents for Site-Directed Fluorescence Labeling allow the incorporation of fluorescent unnatural amino acids into proteins in a site-direct and quantitative fashion. Various fluorescent dyes are available including those for 488 nm, 543 nm and 633 nm excitation.





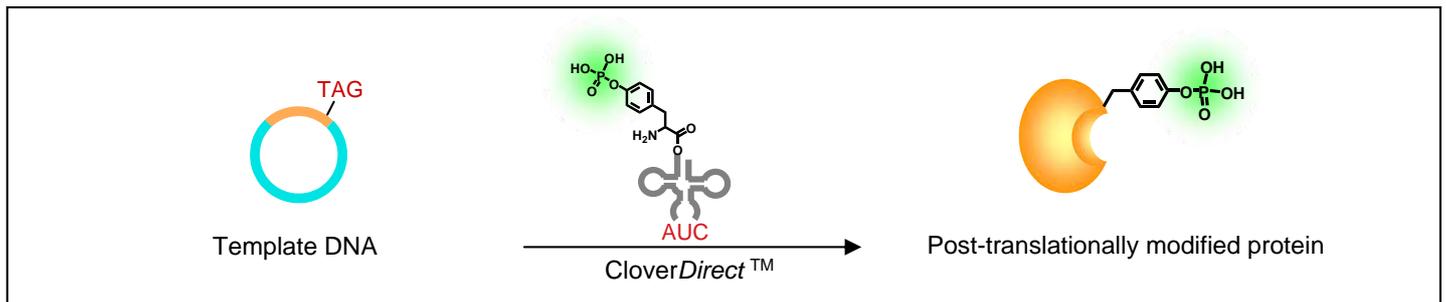
Site-Directed Biotin Labeling

CloverDirect™ tRNA Reagents for Site-Directed Biotin Labeling allow the incorporation of biotinylated unnatural amino acids into proteins in a site-directed and quantitative fashion. Labeling proteins are available for the oriented immobilization onto avidin-coated plates and beads. The biotinylated amino acids have one or two aminohexyl liners between amino acid and biotin.



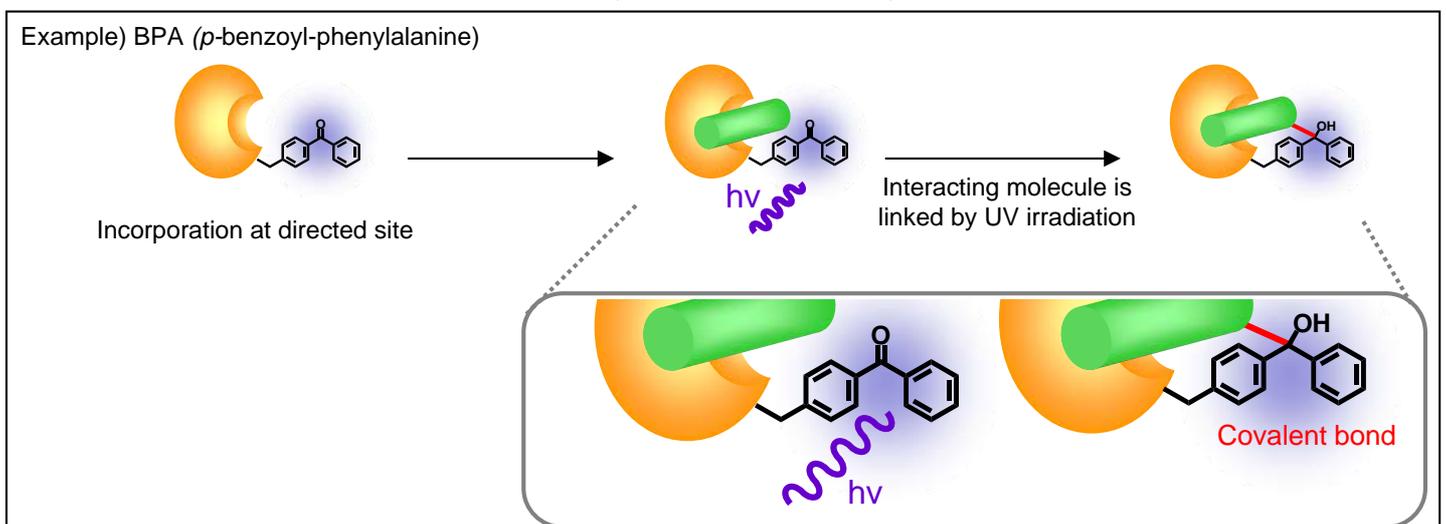
Site-Directed Post-Translational Modification

It is not easy to prepare post-translationally modified proteins (phosphorylation, methylation, etc.). CloverDirect™ tRNA Reagents for Site-Directed Post-Translational Modification allow the incorporation of modified amino acids into proteins in a site-directed and quantitative fashion.



Site-Directed Unnatural Mutagenesis

By incorporation of unnatural amino acids containing functional groups, novel functional proteins can be designed and synthesized. CloverDirect™ tRNA Reagents for Site-Directed Unnatural Mutagenesis allow the incorporation of unnatural amino acids with PEG, photo-crosslinking, photo-isomerizable groups, etc.





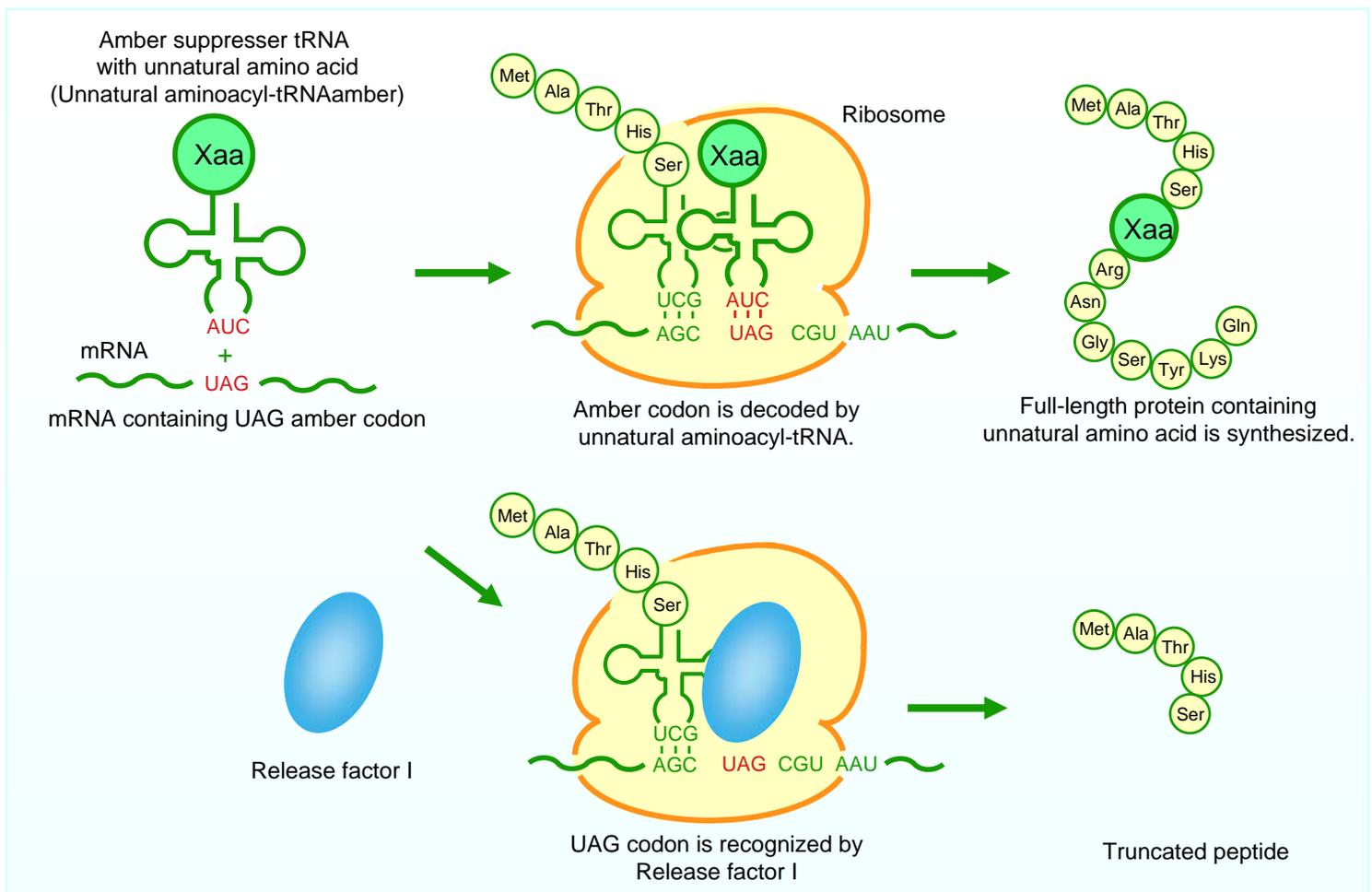
Product Description

[Principle of incorporation of unnatural amino acids]

Incorporation position of unnatural amino acids is defined by a UAG amber codon or CGGG four-base codon. An unnatural aminoacyl-tRNA recognizes the UAG amber codon or the CGGG codon during translation. Consequently, the unnatural amino acid is incorporated at the directed site of the protein. By using two tRNAs for amber and four-base codons, dual-labeled proteins can be obtained which are available for fluorescence resonance energy transfer (FRET).

[UAG amber codon]

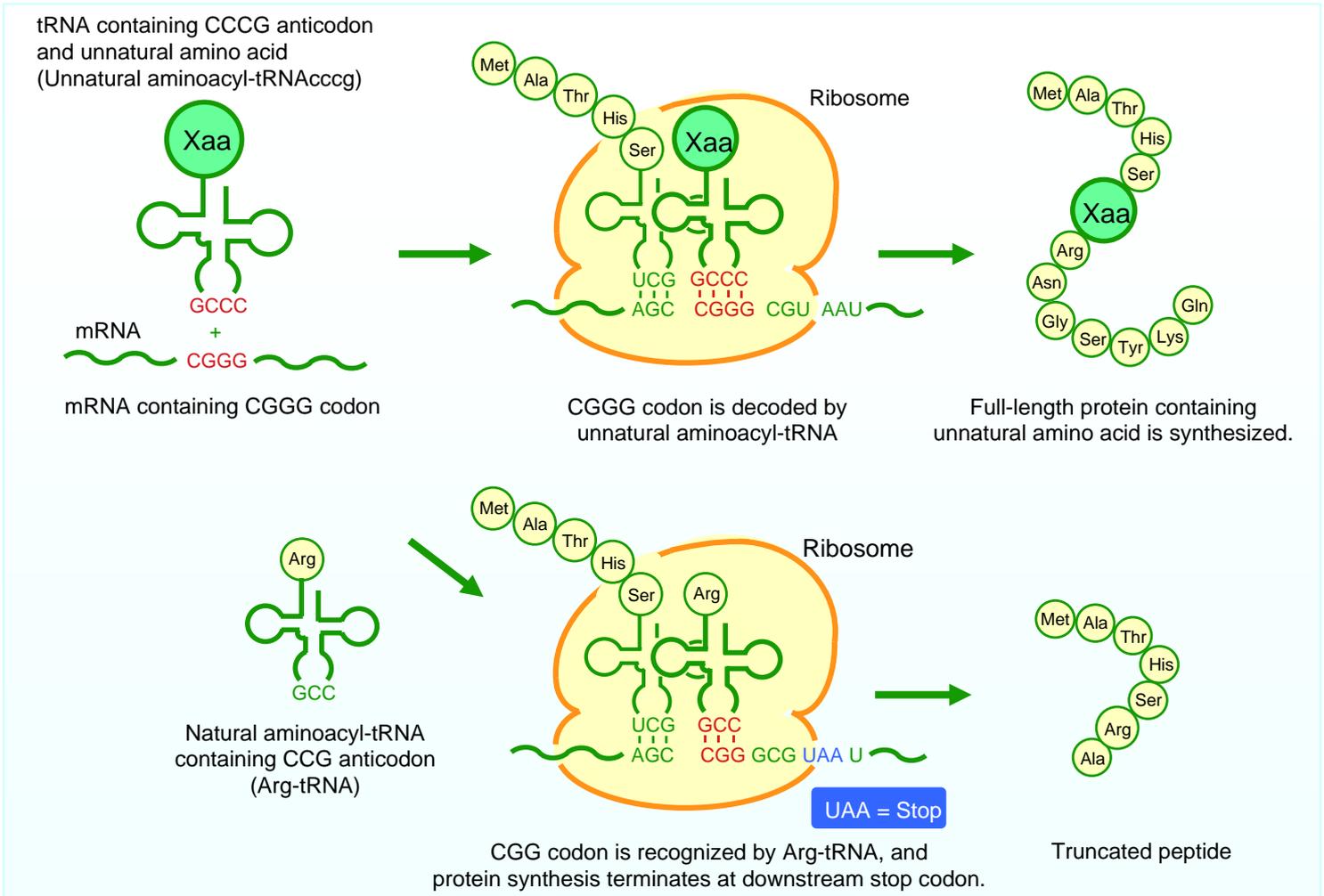
If the UAG codon is recognized by the amber suppressor tRNA, full-length protein containing the unnatural amino acid is successfully synthesized. On the contrary, if the UAG codon is recognized by release factor 1 (RF1) which is one of the termination factors, the protein synthesis is terminated. Therefore, the translation product obtained as a full-length protein contains the unnatural amino acid at 100% efficiency.





[CGGG four-base codon]

If the CGGG codon is recognized by the four-base anticodon tRNA, full-length protein containing the unnatural amino acid is successfully synthesized. On the contrary, if the CGG is recognized as a triplet codon by Arg-tRNA, the reading frame shifts to +1 frame and a downstream stop codon terminates the protein synthesis. Therefore, the translation product obtained as a full-length protein contains the unnatural amino acid at 100% efficiency.



Incorporation at N-terminal regions (within 20 amino acid residues from the N-terminus) in response to CGGG codon sometimes results in the production of full-length proteins without unnatural amino acids, possibly because of spontaneous +1 frameshifting. In such case, ProteinExpress recommend the use of ProX™ tag, which is original peptide tag developed for the CGGG codon-mediated incorporation of unnatural amino acids.

5'-	AUG	UCU	AAA	CAA	AUC	GAA	GUA	AAC	CGGG	UCU	AAU	GAG	-3'
	Met	Ser	Lys	Gln	Ile	Glu	Val	Asn	Xaa	Ser	Asn	Glu	

Sequence of ProX™ tag



Product Description

[Product Contents]

- | | |
|--|-----|
| · Unnatural aminoacyl-tRNA (See note1) | X 1 |
| · tRNA dissolving buffer | X 1 |

Note 1 : One tube contains unnatural aminoacyl-tRNA sufficient for 300 μ L of *in vitro* translation reaction. Once thawed, unnatural aminoacyl-tRNA can be stored at -70 °C for 2 months.

[Equipment and reagents to be supplied by others]

Protein Expression

- Cell-free translation system(*E.coli*)
(e.g. RTS100 *E.coli* HY kit; Roche Applied Science, #3186156)
- Expression gene containing UAG or CGGG codon (plasmid DNA, linear DNA, or messenger RNA).
(Suitable expression vector should be used for the cell-free translation system.)

Purification, buffer exchange, and concentration

- Affinity column and buffer for purification
(e.g. His SpinTrap™ kit; GE Healthcare Science , #28-9321-71)
- Ultrafiltration membrane and buffer
(e.g. ULTRAFREE®-0.5 Centrifugal Filter Devices 10k; Millipore , UFV5BC00)



[Brief protocol]

Step 1 Protein Expression

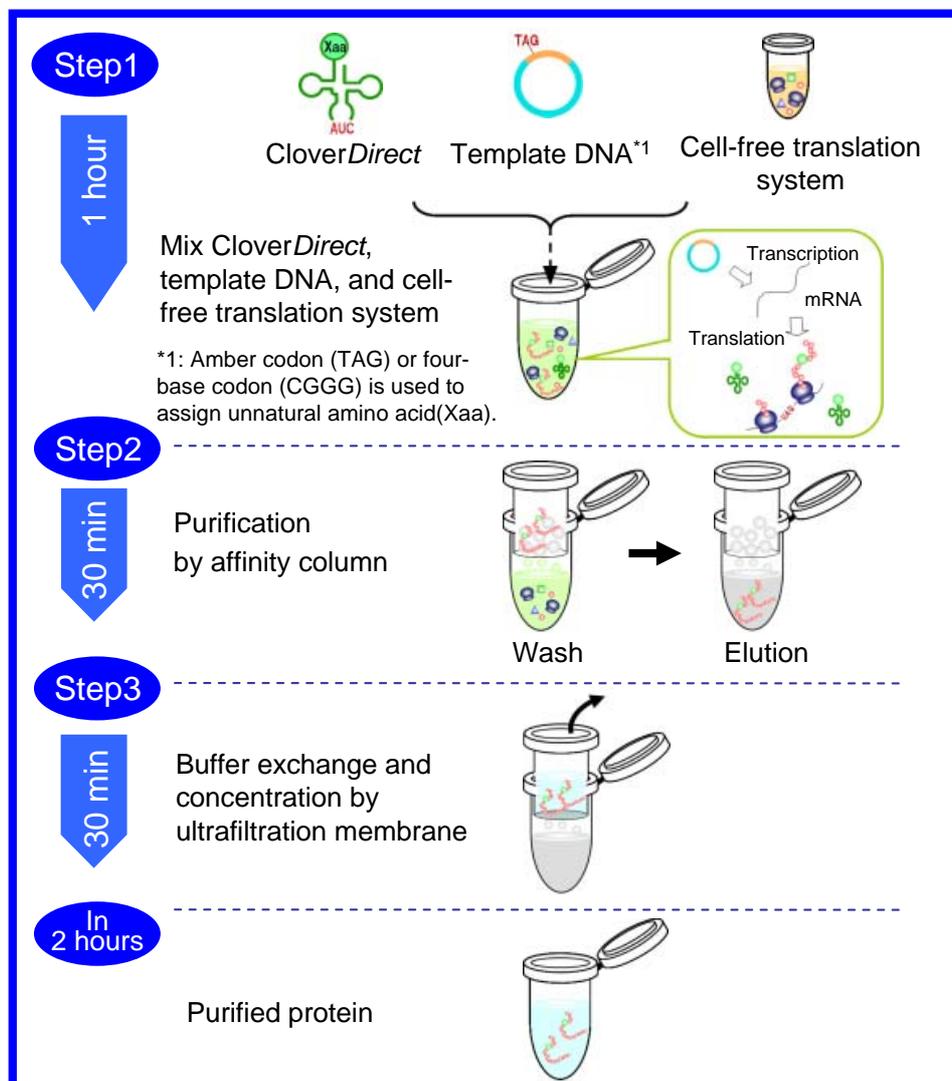
Dissolve unnatural aminoacyl-tRNA with tRNA buffer, and mix with template DNA and cell-free translation system. The mixture is incubated for 1 hour to synthesize protein containing unnatural amino acid.

Step 2 Purification

Reaction mixture includes several proteins derived from cell-free translation system and unnatural amino acid which is not incorporated into protein. Full-length protein containing unnatural amino acid can be isolated by purification for C-terminal tag such as His tag.

Step 3 Buffer exchange and concentration (optional)

Buffer exchange and concentration can be done by ultrafiltration membrane. Purified protein containing unnatural amino acid can be directly used for the downstream experiment.



Technical Overview of CloverDirect™



Product Description

[Points to note]

1. Protein expression

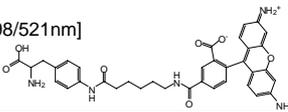
Confirm your protein can be expressed in *E.coli* cell-free translation system.

In case of very low expression of a wild-type gene that does not contain UAG codon or CGGG codon, optimization of nucleotide sequence (codon usage, addition of N-terminal tags, etc.) is required to improve the expression.

ProteinExpress provides custom service for protein expression using CloverDirect™ including gene construction for efficient expression.

2. Incorporation site-dependency

Some unnatural amino acids are allowed to be incorporated only at the N-terminal region (within 20 amino acid residues from the N-terminus). Please check the product list for details.

Pin-point Fluorescence Labeling	Site-dependency (*1)		codon	Q'ty (translation)	Product No
	N-terminal	Internal, C-terminal			
CR110-X-AF [5-CR110-X : Abs/Em = 498/521nm] 	○	×	Amber	300 μL	CLD1001
	●	●	CGGG	5 X 300 μL	CLD2001
	●	●	CGGG	5 X 300 μL	CLD2002

○ : Available
 △ : Available in some sites
 × : Unavailable

N-terminal region = within 20 amino acid residues from the N-terminus

Incorporation at N-terminal regions (within 20 amino acid residues from the N-terminus) in response to CGGG codon sometimes results in the production of full-length proteins without unnatural amino acids, possibly because of spontaneous +1 frameshifting. In such case, ProteinExpress recommend the use of ProX™ tag, which is original peptide tag developed for the CGGG codon-mediated incorporation of unnatural amino acids.

5'-	AUG	UCU	AAA	CAA	AUC	GAA	GUA	AAC	CGGG	UCU	AAU	GAG	-3'
	Met	Ser	Lys	Gln	Ile	Glu	Val	Asn	Xaa	Ser	Asn	Glu	

Sequence of ProX™ tag

3. Custom Services

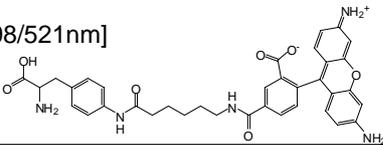
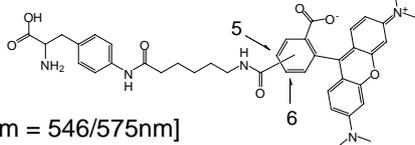
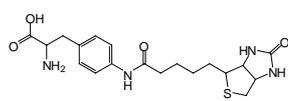
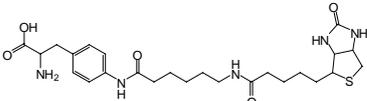
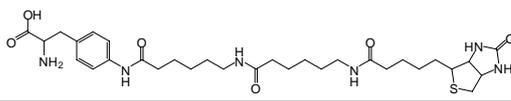
ProteinExpress provides custom services for the synthesis of unnatural aminoacyl-tRNAs (fluorescent-labeled, functional amino acids, etc.), which allows the expression of proteins with your requested unnatural amino acids.

- Custom Services for Unnatural Aminoacyl-tRNA.....p.11

ProteinExpress also provides custom services for the expression of proteins with unnatural amino acids including construction of recombinant DNA, cell-free translation, and purification of proteins.

- Custom Service for Protein Expression using CloverDirect™.....p.11



Site-Directed Fluorescence Labeling	Site-dependency (*1)		codon	Q'ty (translation)	Product No
	N-terminal	Internal, C-terminal			
CR110-X-AF [5-CR110-X : Abs/Em = 498/521nm] 	○	×	Amber	300 μL	CLD1001
			CGGG	5 X 300 μL	CLD2001
				5 X 300 μL	CLD2002
HiLyte Fluor™ 488-AF [HiLyte Fluor™ 488 : Abs/Em = 497/525nm] Not Available	○	×	Amber	300 μL	CLD01
			CGGG	5 X 300 μL	CLD05
				5 X 300 μL	CLD2004
TAMRA-X-AF  [5(6)-TAMRA-X : Abs/Em = 546/575nm]	○	×	Amber	300 μL	CLD02
			CGGG	5 X 300 μL	CLD06
				Contact to OLYMPUS	
ATTO633-AF [ATTO633 : Abs/Em = 629/657nm] Not Available	○	×	Amber	300 μL	CLD03
			CGGG	5 X 300 μL	CLD07
				5 X 300 μL	CLD2008
ATTO655-X-AF [ATTO655-X : Abs/Em = 633/684nm] Not Available	○	×	Amber	300 μL	CLD1009
			CGGG	5 X 300 μL	CLD2009
				300 μL	CLD1010
				5 X 300 μL	CLD2010
Site-Directed Biotin Labeling					
Biotin-AF [Biotin] 	○	○	Amber	5 X 300 μL	CLD2101
			CGGG	5 X 300 μL	CLD2102
Biotin-X-AF [Biotin-X] 	○	○	Amber	5 X 300 μL	CLD2103
			CGGG	5 X 300 μL	CLD2104
Biotin-XX-AF [Biotin-XX] 	○	△	Amber	300 μL	CLD04
			CGGG	5 X 300 μL	CLD08
				5 X 300 μL	CLD2106

*1 : Incorporation site-dependency

[Please inquire about the price]

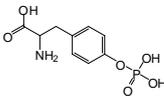
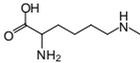
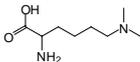
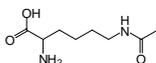
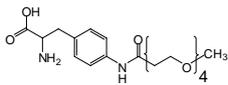
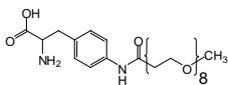
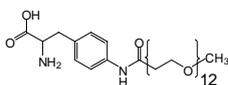
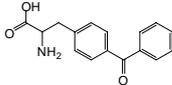
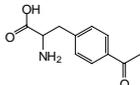
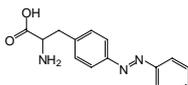
Some unnatural amino acids are incorporated in a incorporation site-dependent manner.
 (N-terminal region = within 20 amino acid residues from the N-terminus.)

○ : Available

△ : Available in some sites

× : Unavailable



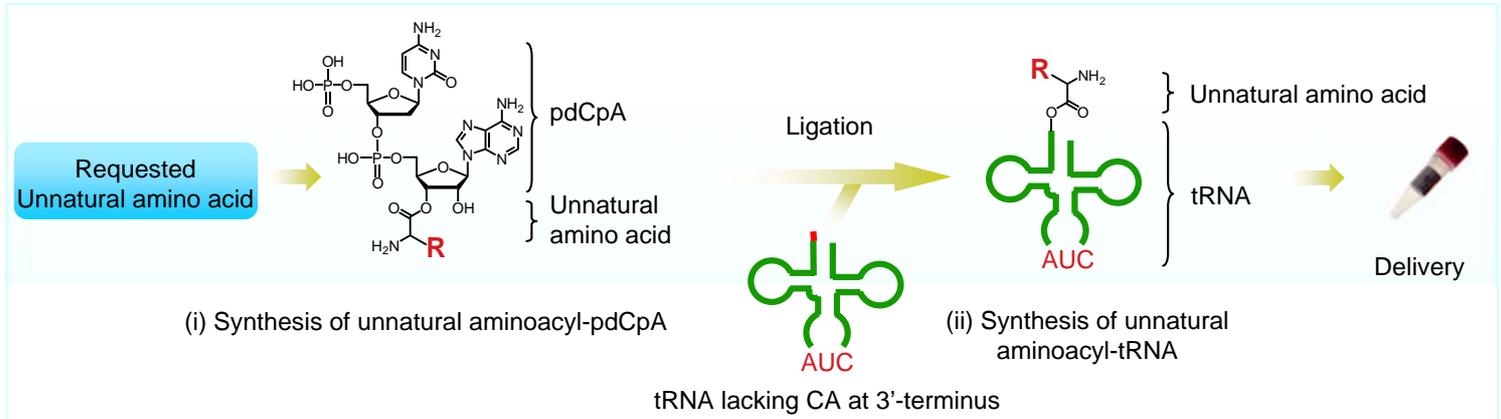
Site-Directed Post-Translational Modification	Incorporable site (*1)		codon	Q'ty (translation)	Product No
	N-terminal	Internal, C-terminal			
Tyr(PO₃H₂) [O-phospho-Tyr] 	○	△	Amber	5 X 300 μL	CLD2201
			CGGG	5 X 300 μL	CLD2202
Lys(Me) [ε-methyl-Lys] 	○	○	Amber	5 X 300 μL	CLD2203
			CGGG	5 X 300 μL	CLD2204
Lys(Me₂) [ε-dimethyl-Lys] 	○	○	Amber	5 X 300 μL	CLD2205
			CGGG	5 X 300 μL	CLD2206
Lys(Ac) [ε-acetyl-Lys] 	○	○	Amber	5 X 300 μL	CLD2207
			CGGG	5 X 300 μL	CLD2208
Site-Directed Unnatural Mutagenesis					
PEGylated amino acids					
PEG4-AF [Methyl-PEG4] 	○	○	Amber	5 X 300 μL	CLD2301
			CGGG	5 X 300 μL	CLD2302
PEG8-AF [Methyl-PEG8] 	○	○	Amber	5 X 300 μL	CLD2303
			CGGG	5 X 300 μL	CLD2304
PEG12-AF [Methyl-PEG12] 	○	△	Amber	5 X 300 μL	CLD2305
			CGGG	5 X 300 μL	CLD2306
Cross-linking amino acids					
BPA [ρ-benzoyl-phenylalanine] 	○	○	Amber	5 X 300 μL	CLD2321
			CGGG	5 X 300 μL	CLD2322
AcPhe [ρ-acetyl-phenylalanine] 	○	○	Amber	5 X 300 μL	CLD2323
			CGGG	5 X 300 μL	CLD2324
Photo-isomerizable amino acid					
azoAla [ρ-phenylazophenyl-alanine] 	○	○	Amber	5 X 300 μL	CLD2331
			CGGG	5 X 300 μL	CLD2332

[Please inquire about the price]



[Custom Services for Unnatural Aminoacyl-tRNA]

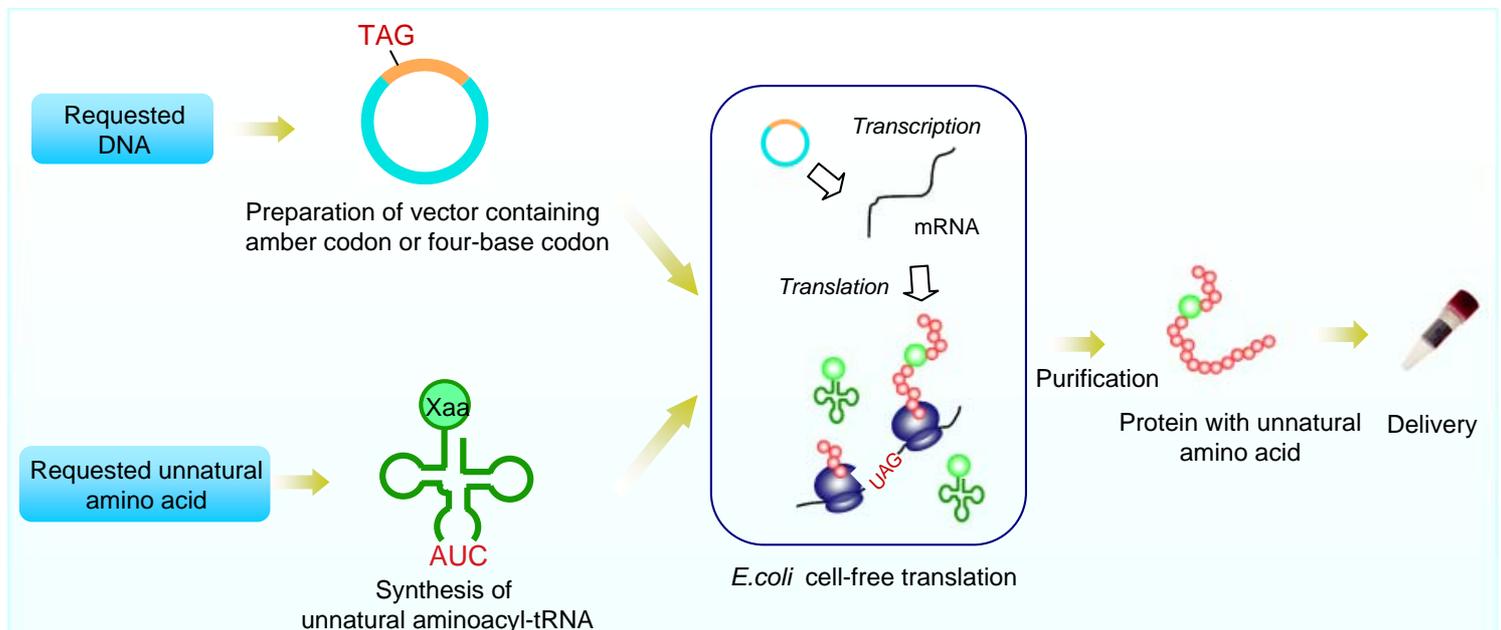
ProteinExpress provides custom services for the synthesis of unnatural aminoacyl-tRNAs (fluorescent-labeled, functional amino acids, etc.), which allows the expression of proteins with your requested unnatural amino acids.



Custom Synthesis of Unnatural Aminoacyl-tRNA

[Custom Service for Proteins with Unnatural Amino Acids]

ProteinExpress provides custom services for the expression of proteins with your requested unnatural amino acids at requested positions, including artificial gene synthesis, cell-free translation, and protein purification.



Custom Service for Protein with Unnatural Amino Acids

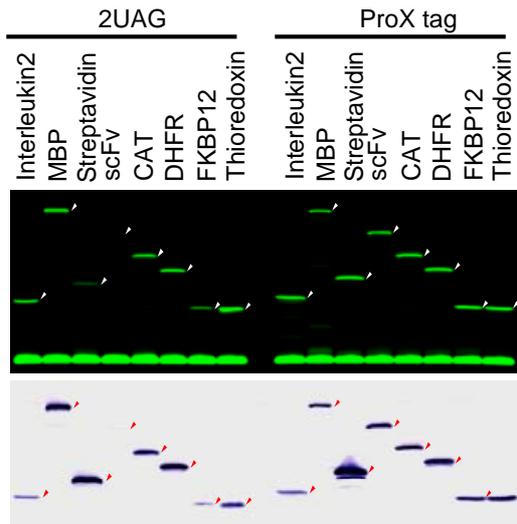


[Expression of site-directly labeled proteins]

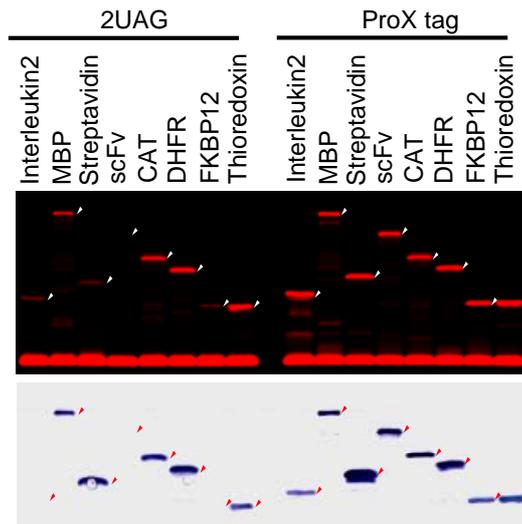
Fluorescent- and biotin-labeled unnatural amino acids are incorporated into eight prokaryote and eukaryote proteins. The site-directly fluorescent-labeled proteins can be visualized on SDS-PAGE using a laser-based fluorescence scanner. The proteins are also detectable by an antibody against tag peptide or biotin. A 0.25 ~ 1 μ L of translational reaction mixture is sufficient for the detection.

· Labeled unnatural amino acids that are not incorporated into proteins are detected at the bottom of SDS-PAGE gel.

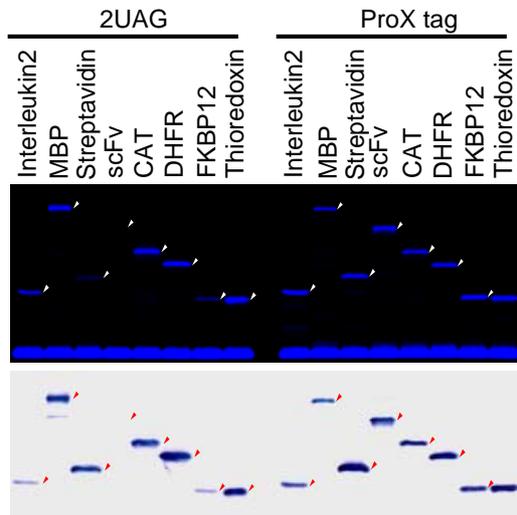
HiLyte Fluor 488



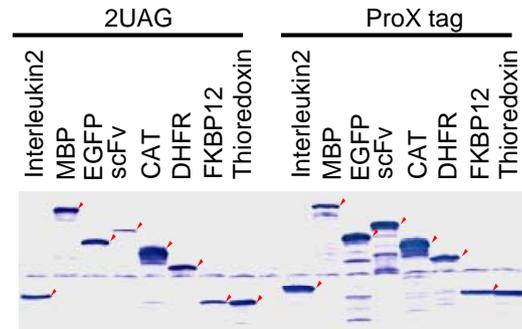
TAMRA



ATTO633



Biotin



2UAG : UAG codon is inserted after initiator AUG codon.

ProX tag : ProX tag is fused to the N-terminus.

Applied volume: 0.25 μ L of translational reaction mix

Fluorescence image (Top) are visualized with Ex and Em wavelengths listed below:

HiLyteFluor488 Ex : 488nm / Em : 520 nm

TAMRA Ex : 532nm / Em : 580 nm

ATTO633 Ex : 635nm / Em : 670 nm

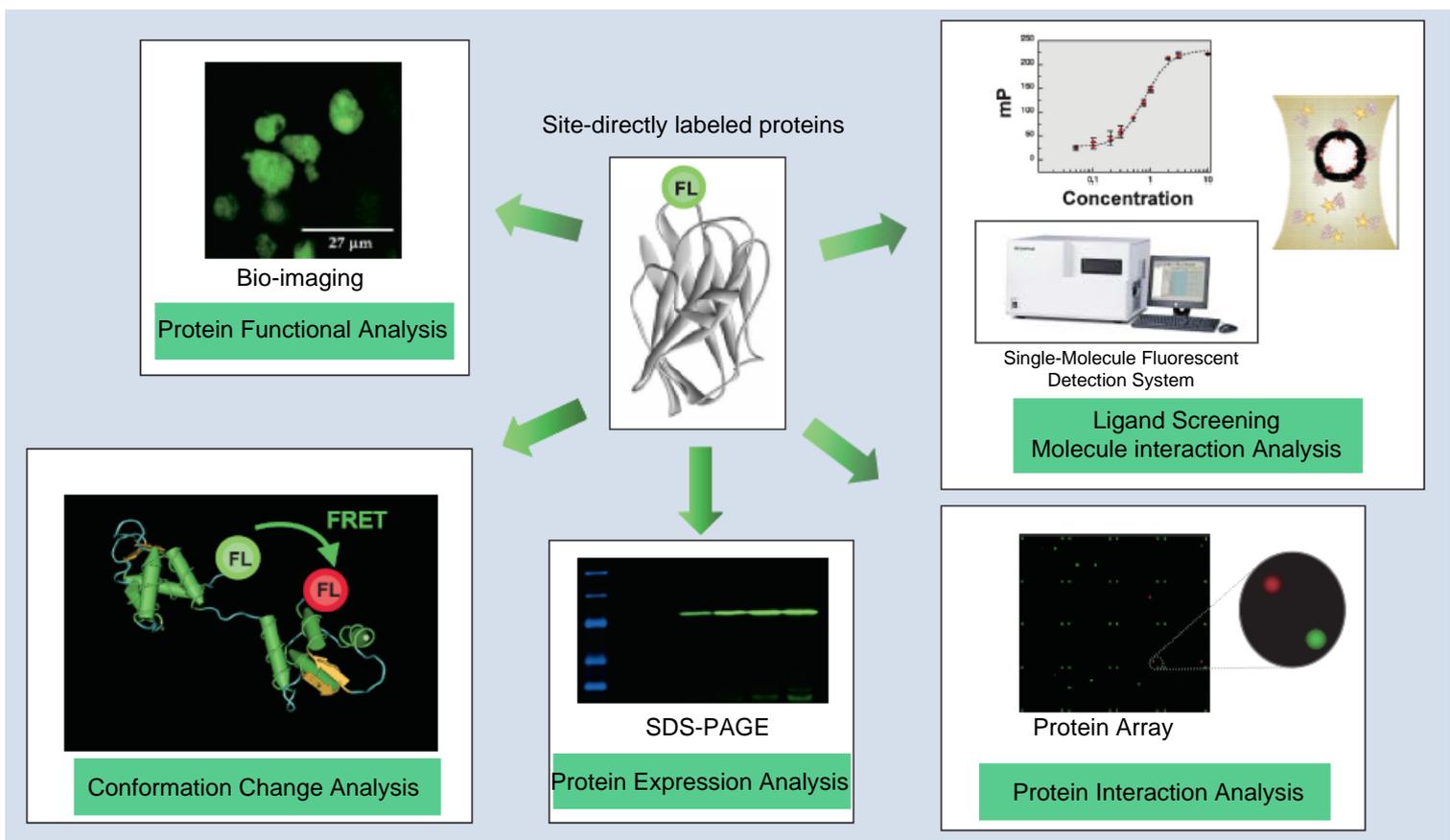
Western blotting (Bottom) are visualized by anti-His tag antibody (for fluorescent amino acids) and anti-biotin antibody (for biotin).



[Applications for site-directly fluorescent labeled proteins]

The site-directly fluorescent-labeled proteins are available to the following measurements.

- Interaction analysis using single molecule fluorescence analysis
Single Molecule fluorescence detection system (MF20 / Fluor Point-Light MF20; OLYMPUS)
- Conformation analysis of protein by inter- or intra-molecular fluorescence resonance energy transfer (FRET)
- Functional analysis in cell imaging
- Interaction analysis in protein array
- Expression analysis by in-gel fluorescent detection of SDS-PAGE





References / Questions about Products

[References]

- 1) FRET analysis of protein conformational change through position-specific incorporation of fluorescent amino acids
Daisuke Kajihara, Ryoji Abe, Issei Iijima, Chie Komiyama, Masahiko Sisido, Takahiro Hohsaka
Nature Methods., 3, 923-929 (2006).
- 2) Position-specific incorporation of biotinylated non-natural amino acids into a protein in a cell-free translation system
Takayoshi Watanabe, Norihito Muranaka, Issei Iijima, Takahiro Hohsaka
Biochem. Biophys. Res. Commun., 361, 794-799 (2007)
- 3) Comprehensive screening of amber suppressor tRNAs suitable for incorporation of non-natural amino acids in a cell-free translation system
Hikaru Taira, Yosuke Matsushita, Kenji Kojima, Kaori Shiraga, Takahiro Hohsaka
Biochem. Biophys. Res. Commun., 374, 304-308 (2008).
- 4) Efficient Incorporation of Nonnatural Amino Acids with Large Aromatic Groups into Streptavidin in In Vitro Protein Synthesizing Systems
Takahiro Hohsaka, Daisuke Kajihara, Yuki Ashizuka, Hiroshi Murakami, Masahiko Sisido
J. Am. Chem. Soc., 121, 34-40 (1999).

[Questions about Products]

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*CloverDirect*TM

tRNA Reagents for Site-Directed Protein Functionalization

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ProteinExpress

Catalogue 2009/09