

Anti-p62 (SQSTM1) (Human) mAb -Alexa Fluor[®] 647

CODE No. M162-A64

CLONALITY Monoclonal
CLONE 5F2
ISOTYPE Mouse IgG1 κ
QUANTITY 100 μ L, 1 mg/mL

SOURCE Purified IgG from hybridoma supernatant
IMMUNOGEN Recombinant Human p62 (120-440 a.a.)
FORMULATION PBS containing 1% BSA and 0.1% ProClin 150
STORAGE This antibody solution is stable for one year from the date of purchase when stored at 4°C.

APPLICATIONS-CONFIRMED

Immunocytochemistry 5 μ g/mL
Flow cytometry 1 μ g/mL

SPECIES CROSS REACTIVITY on IC

Species	Human	Mouse	Rat	Hamster
Cells	Transfectant	Not tested	Not tested	Not tested
Reactivity	+			

Entrez Gene ID 8878 (Human)

REFERENCES
1) Ichimura, Y., *et al.*, *J. Biol. Chem.* **283**, 22847-22857 (2008)
2) Komatsu, M., *et al.*, *Cell* **131**, 1149-1163 (2007)

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Immunocytochemistry

- 1) Spread the cells in the nutrient condition on a glass slide, then incubate in a CO₂ incubator overnight.
- 2) Remove the culture supernatant by careful aspiration.
- 3) Fix the cells by immersing the slide in 4% paraformaldehyde /PBS for 10 minutes at room temperature (20~25°C).
- 4) Prepare a wash container such as a 500 mL beaker with a magnetic stirrer. Then wash the fixed cells on the glass slide by soaking the slide with a plenty of PBS in the wash container for 5 minutes. Take care not to touch the cells. Repeat another wash once more.
- 5) Immerse the slide in 100 µg/mL digitonin in PBS for 10 minutes at room temperature.
- 6) Wash the slide in a plenty of PBS as in the step 4).
- 7) Add 200 µL of Clear Back (human Fc receptor blocking reagent, MBL, code no. MTG-001) onto the cells and incubate for 5 minutes at room temperature.
- 8) Add 200 µL of the primary antibody diluted with 2% fetal calf serum (FCS)/PBS as suggested in the **APPLICATIONS** onto the cells and incubate for 60 minutes at room temperature. (Optimization of antibody concentration or incubation condition is recommended if necessary.)
- 9) Wash the slide in a plenty of PBS as in the step 4).
- 10) Promptly add mounting medium onto the slide, then put a cover slip on it.

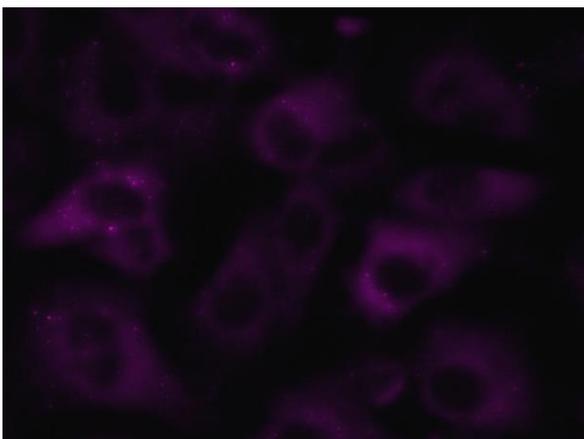
(Positive control for Immunocytochemistry; A549)



Immunocytochemical detection of p62 in A549

Upper: Starved A549

Lower: Nutrient A549



Flow cytometric analysis for adherent cells

- 1) Detach the cells from culture dish.
- 2) Wash the cells once with 1 mL of washing buffer [PBS containing 2% fetal calf serum (FCS)].
- 3) Add 200 μ L of 4% paraformaldehyde to the cell pellet after tapping. Mix well, then fix the cells for 10 minutes at room temperature.
- 4) Wash the cells twice with 1 mL of washing buffer.
- 5) Add 200 μ L of 100 μ g/mL digitonin in PBS to the cell pellet after tapping. Mix well, then permeabilize the cells for 10 minutes at room temperature.
- 6) Wash the cells once with 1 mL of washing buffer.
- 7) Resuspend the cells with washing buffer (5×10^6 cells/mL).
- 8) Add 100 μ L of the cell suspension into each tube, and centrifuge at 500 x g for 1 minute at room temperature (20~25°C). Remove supernatant by careful aspiration.
- 9) Add 20 μ L of Clear Back (human Fc receptor blocking reagent, MBL, code no. MTG-001) to the cell pellet after tapping. Mix well and incubate for 5 minutes at room temperature.
- 10) Add 40 μ L of the primary antibody at the concentration as suggested in the **APPLICATIONS** diluted in the washing buffer. Mix well and incubate for 30 minutes at room temperature.
- 11) Add 1 mL of the washing buffer followed by centrifugation at 500 x g for 1 minute at room temperature. Remove supernatant by careful aspiration. Repeat another wash once more.
- 12) Resuspend the cells with 500 μ L of the washing buffer and analyze by a flow cytometer.

(Positive control for Flow cytometry; A549)

