

Hexokinase (HK, EC 2.7.1.1) is an enzyme that phosphorylates hexoses (six-carbon sugars), forming hexose phosphate. In most organisms, glucose is the most important substrate of hexokinases, and glucose-6-phosphate is the most important product. Hexokinase can transfer an inorganic phosphate group from ATP to a substrate.

Hexokinases should not be confused with glucokinase, which is a specific isoform of hexokinase. While other hexokinases are capable of phosphorylating several hexoses, glucokinase acts with a 50-fold lower substrate affinity and its only hexose substrate is glucose.

The colorimetric assay for serum glucose previously reported was modified by replacing the light-sensitive phenazine methosulfate with diaphorase resulting in a simplified, one-reagent system. The sensitivity, specificity, and regeneration of NAD originally reported were maintained. The following reaction scheme summarizes the modified glucose assay:

D-Glucose+ATP<u>HK</u>→D-Glucose-6-phosphate+ADP

D-Glucose-6-phosphate+NAD+ $\xrightarrow{G-6-PDH}$ Glucono- δ -lactone-6-phosphate+NADH+H+

The increased formation of NADH is in absorbance at 340 nm. The addition is directly proportional to the amount of glucose in the sample.

Hexokinase (HK)

A new generation of hexokinase, which was recently produced by CUSAg, makes possible the development of biochemistry assay. The enzyme can be used for serum glucose assay, which is a simple, rapid, sensitive and homogeneous assay that can be performed using automated systems for high-throughput assays.

Properties	Specification
Appearance	White amorphous powder, lyophilized
Activity	≥100 U/mg -solid or more
Genus	Microorganism
Purification method, Purity	Affinity chromatography, ion exchange, $>$ 90% (SDS-PAGE)
Stability	Store at -20°C
Optimum pH	8.0-8.5
Optimum temperature	45°C
pH Stability	pH 8.0-9.0 (25°C, 20 h)
Thermal stability	≤40°C (pH 7.0, 30 min)
Inhibitors	Metal ions and NaN3, etc
Catalog Number	CSB-DE001

1 Thermal stability

⊚ Thermal stability of HK-solid: Three batches of HK-solid were stored at -20°C and 37°C for 14 days, respectively. During this period, enzyme activity of HK was determined on our biochemistry platform. Fig.1 shows that both HK residual activities are parallel at different temperatures, and the relative bias was <10%, representing that our CUSAg enzyme was stable at -20°C and 37°C.

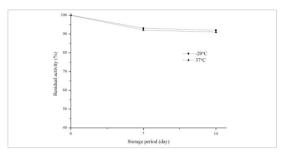


Fig.1 Thermal stability of HK-solid

© Thermal stability of HK-reagent: CUSAg in-house serum glucose reagents prepared with our hexokinase were respectively stored at 4°C and 37°C for 14 days. During the incubation time, the serum glucose control was tested at the interval of 7 days. Fig.2 shows that the relative bias was <10%, representing that our glucose reagent was stable invariably at 4°C or 37°C stored.

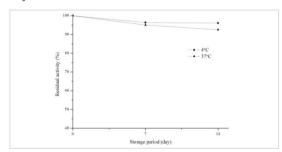


Fig.2 Thermal stability of HK-reagent

2 Clinical comparison

Our CUSAg glucose reagent was also evaluated in medium-scale clinical trials with random blood samples from donations (n=80). Fig.3 shows that the correlation coefficient (r) is as high as 0.99 between in-house biochemistry assay and commercial assay. These results show good agreement between the two systems.

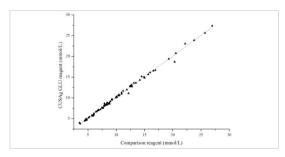


Fig.3 Clinical comparison of in-house HK-reagent and commercial kit

Accuracy

Two levels of glucose control was analyzed in replicates of three using our two lots of reagents on the CUSAg biochemistry platform. Data from this study are summarized in the below table.1, the relative bias was <6%.

Table.1 Accuracy of glucose reagent

Control	Control Target value (mmol/L)		rmined mmol/L		Mean Con. (mmol/L)	Bias (%)
1	6.11	5.75	5.83	5.80	5.79	-5.2
2	15.60	14.82	14.94	14.81	14.86	-4.8

Precision

Two members of serum samples based panel were assayed, using a single lot of reagents, in replicates of ten on the CUSAg biochemistry platform (from 3.89 to 6.11 mmol/L as normal). As shown in table.2, the system shows excellent precision with $CV \le 1\%$.

Table.2 Precision profile of glucose reagent

Panel Member	n	Mean Con.(mmol/L)	SD	%CV
1	10	4.81	0.05	0.9
2	10	15.39	0.13	0.9

6 Enzymatic properties

Optimum pH: Our HK enzyme activity was determined at 30°C in different buffer (50 mM buffer: pH 6.0-7.5 PIPES-NaOH; pH 8.0-9.0 Tris-HCl; pH 9.5-11.0 Glycine-NaOH). As shown in Fig.4, the optimum pH for HK is from 8.0 to 8.5.

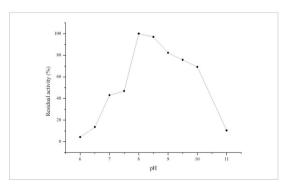


Fig.4 Optimum pH

(30°C in the 50mM buffer solution: pH 6.0-7.5 PIPES-NaOH; pH 8.0-9.0 Tris-HCl; pH 9.5-11.0 Glycine-NaOH)



Optimum temperature: Our HK enzyme activity was determined at different temperatures in 50 mM Tris-HCl buffer. Fig.5 shows that the optimum temperature of HK is 45°C.

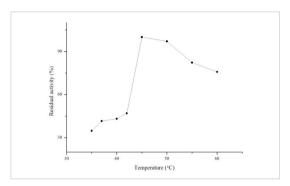


Fig.5 Optimum temperature

(50mM Tris-HCl buffer, pH 8.0)

© pH Stability: 10 U/mL of CUSAg HK was incubated at 25°C for 20 hours in different buffer (100 mM buffer: pH 4.0-6.0 Acetate-NaOH; pH 6.5-7.5 K-phosphate; pH 8.0-9.0 Tris-HCl; pH 9.5-11.0 Glycine-NaOH), and the residual activities were determined with biochemistry assay. As shown in Fig.6, HK is stable from pH 8.0 to 9.0.

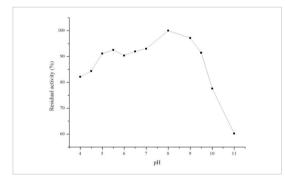


Fig.6 pH stability

(25°C,20 h-treatment in the 100m M buffer solution: pH 4.0-6.0 Acetate-NaOH; pH 6.5-7.5 K-phosphate; pH 8.0-9.0 Tris-HCl; pH 9.5-11.0 Glycine-NaOH; enzyme concentration: 10 U/mL)

● **Thermal stability:** Our CUSAg HK was incubated from 25°C to 55°C for 30 minutes in 50 mM K-phosphate buffer (pH 7.0), containing 5 U/mL of HK and 0.1% BSA, and the residual activities were determined with colorimetric assay on biochemical analyzer. As shown in Fig.7, HK is stable below 40°C.

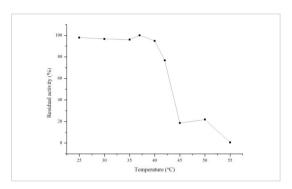


Fig.7 Thermal stability

(30 min-treatment with 50mM K-phosphate buffer, pH 7.0, containing 0.1% BSA, enzyme concentration: 5 U/mL)

© Effect of Various Chemicals: Our CUSAg HK was dissolved at a concentration of 5U/mL in 50 mM, pH 6.5, K-phosphate buffer, which was incubated with each chemical at 30°C for 1h. The activities were determined with biochemistry assay. As shown in table.2, HK can be inhibited by metal ions and NaN₃.

Table.3 Effect of Various Chemicals on HK

Chemical	Concn.(mM)	Residual activity (%)
None	_	100
NaCl	2.0	71
$CuSO_4$	2.0	63
CaCl ₂	2.0	56
KCI	2.0	56
NH₄CI	2.0	56
$MnCl_2$	2.0	50
$FeCl_3$	2.0	29
$MgSO_{\scriptscriptstyle{4}}$	2.0	47
NiCl ₂	2.0	35
NaN_3	20.0	42



ADDR: No.818 Gaoxin Avenue, Wuhan Hi-tech Medical Devices Park, Donghu High-tech Development Zone 430206, Wuhan City, Hubei Province, P.R. China.

TEL: +86-27-87196282 Ext.837/853 **FAX:** +86-27-87196150

EMAIL:cusag@cusag.cn **WEB:**

WEB: www.cusag.org