



KB01010
Nitrite/Nitrate
Assay Kit

96 well plate
100/200/400 tests

Table of contents

| | | |
|-----|--|----|
| 1. | General information | 1 |
| 2. | Technical specifications | 2 |
| 3. | Materials and storage | 3 |
| 4. | Introduction | 4 |
| 5. | Assay principle | 5 |
| 6. | Assay preparation | 6 |
| 7. | Sample preparation | 8 |
| 8. | Assay protocol | 9 |
| 9. | Data analysis | 11 |
| 10. | Troubleshooting | 13 |
| 11. | Additional information | 15 |
| 12. | Related products | 15 |
| 13. | Warranties and limitation of liability | 16 |

1. General information

PRECAUTIONS

Please read this manual carefully before beginning the assay.

This product is designed for **research use only**. It is not approved for human or animal use or clinical diagnosis. All chemicals should be handled with care and in accordance with laboratory safety practices. It is recommended to use basic Personal Protective Equipment.

Do not use after the expiration date stated on the packaging.

Do not mix or substitute reagents or materials from other kit batches or vendors.

For the **material safety data sheet** (MSDS) please contact us at info@bioquochem.com

TECHNICAL RECOMMENDATIONS

Store reagents as indicated in **Materials and storage** section.

Be sure to keep the bottle capped when not in use.

Let the components reach room temperature (RT) before use.

Immediately before use, gently invert and rotate reagent bottles several times to mix the contents thoroughly.

Avoid foaming or bubbles when mixing or reconstituting components.

Avoid cross contamination of samples or reagents by changing pipette tips between sample, standard and reagent additions.

Be sure to use the optimal microplate for the assay. Flat bottom transparent microplates for UV/VIS applications, and black microplates for fluorescence measurements.

2. Technical specifications

Available sizes

100/200/400 tests

Required sample volume

50 µL/test

Compatible samples

Biological fluids (serum, plasma, urine), tissue homogenates, and cell lysates

Type of detection

Colorimetric (540 nm)

3. Materials and storage

MATERIALS SUPPLIED

| Item | No. Tests | Units | Storage |
|--------------------------------|-----------|-------|---------|
| Reagent A | 100 | 1 | -20 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent B | 100 | 1 | -20 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent C | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent D | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent E | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent F | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Standard | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Transparent 96-Well Microplate | 100 | 1 | RT |
| | 200 | 2 | |
| | 400 | 4 | |

MATERIALS NEEDED BUT NOT SUPPLIED

- Double distilled water (ddH₂O) as Milli-Q Ultrapure Water.
- Labware materials (micropipettes, tubes, stirring/mixing equipment).
- Colorimetric microplate reader – equipped with filter for OD 540 nm.

STORAGE CONDITIONS

On receipt, store kit components as indicated above. Under these conditions, the reagents are stable in the original packaging until the expiration date indicated on the outside of the box. Prepare a fresh set of standards for every use.

4. Introduction

Nitric oxide (NO) is an important signalling molecule that participates in virtually every cellular and organ function in the body. NO is a gaseous free radical with a short half-life in vivo. Therefore, the levels of the more stable NO metabolites, nitrite (NO^{2-}) and nitrate (NO^{3-}), are used for the indirect determination of NO.

Biological NO is mainly synthesized from L-arginine by nitric oxide synthases (NOS). NO at physiologic levels has different beneficial functions (e.g. antimicrobial, antioxidant, neurotransmitter, induces vasodilation, inhibits platelet aggregation and low-density lipoprotein (LDL) oxidation, etc.). In contrast, high levels of NO and its derivatives (e.g. peroxynitrite) are cytotoxic and produces oxidative stress.

Dysregulation of NO signalling pathways is associated with the pathogenesis of cardiovascular disorders and inflammatory diseases.

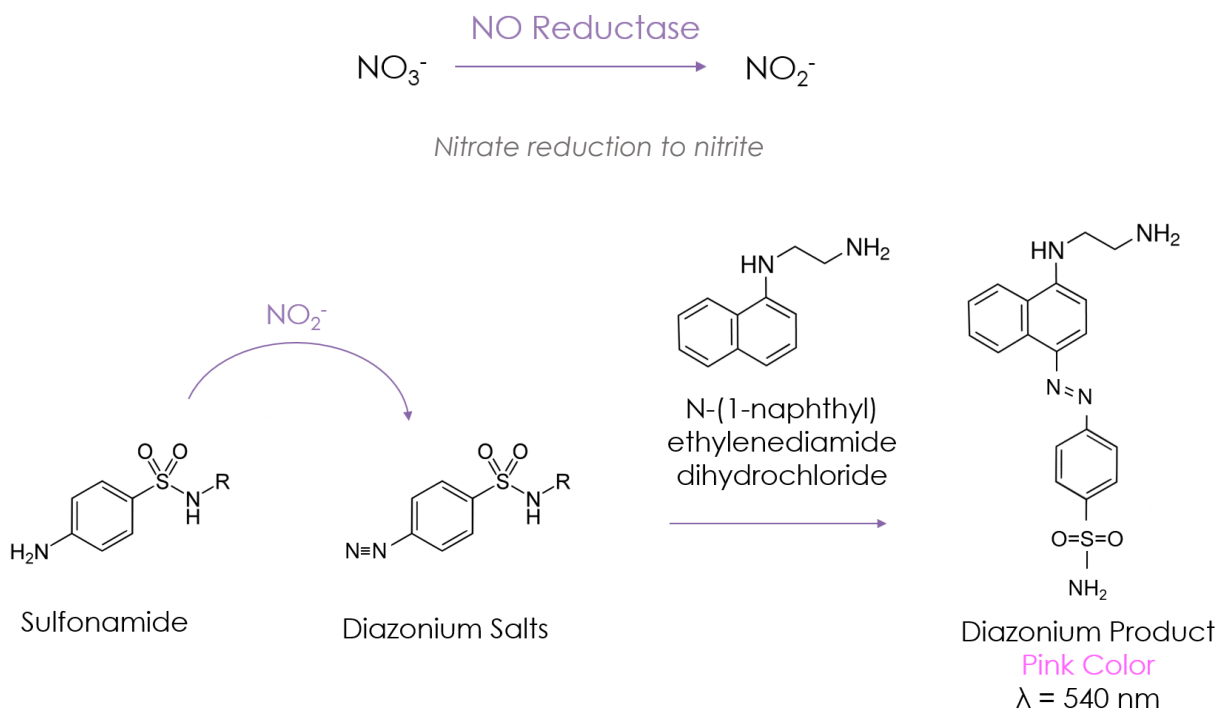
BQC Nitrite/Nitrate Assay Kit is a fast and simple method for the indirect determination of NO in a wide variety of samples.

5. Assay principle

BQC Nitrite/Nitrate Assay Kit is based on the Griess method. Nitrite reacts with Griess reagent to produce an azo compound that can be spectrophotometrically quantified at 540 nm. Nitrate is converted to nitrite by the enzyme nitrate reductase.

Total nitrite (NO_2^-) can be therefore determined following the two step protocol: enzymatic reduction + Griess reaction. Nitrite (NO_2^-) concentration naturally present in the sample can be directly quantify using the Griess reaction. Nitrate (NO_3^-) concentration can be calculated by subtracting nitrite concentration naturally present in the sample from the total nitrite concentration.

Nitrite concentration in a sample is determined from a calibration curve using NO_2^- as standard.



Principle of Nitrite Assay Kit

6. Assay preparation

REAGENT PREPARATION

All assay reagents are ready to use as supplied. Allow the reagents to reach room temperature before use.

R.A. Working Solution: Add 1 mL of ddH₂O to the Reagent A vial and mix well.

ⓘ **CAUTION:** R.A. Working Solution must be freshly prepared and used immediately

R.C. Working Solution: Add 1 mL of ddH₂O to the Reagent C vial and mix well.

ⓘ **CAUTION:** R.C. Working Solution must be freshly prepared and used immediately

STANDARD CALIBRATION

Prepare nitrite (NO₂⁻) standards for the calibration curve from the Standard solution according to the following Table. Prepare the standards immediately prior to each assay. Vortex tubes thoroughly. Discard standard solutions after use.

| Standard | Standard solution (μL) | ddH ₂ O (μL) | Nitrite (μM) |
|--------------------------|------------------------|-------------------------|--------------|
| Std 1 (Reagent Blank) | 0 | 500 | 0 |
| Std 2 | 10 | 490 | 20 |
| Std 3 | 25 | 475 | 50 |
| Std 4 | 50 | 450 | 100 |
| Std 5 | 75 | 425 | 150 |
| Std 6 | 100 | 400 | 200 |

PLATE SET UP

BQC recommends running the standards and samples at least in duplicate (triplicate recommended). There is no specific pattern for using the wells on the plate. A proposed layout of standards (Std) and samples (S) to be measured in duplicate is shown below.

ⓘ **NOTE:** If sample blanks are included in the assay, it is necessary to reserve some wells of the plate for these blanks

| Q | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|-------|-------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | Std 1 | Std 1 | S3 | S3 | S11 | S11 | S19 | S19 | S27 | S27 | S35 | S35 |
| B | Std 2 | Std 2 | S4 | S4 | S12 | S12 | S20 | S20 | S28 | S28 | S36 | S36 |
| C | Std 3 | Std 3 | S5 | S5 | S13 | S13 | S21 | S21 | S29 | S29 | S37 | S37 |
| D | Std 4 | Std 4 | S6 | S6 | S14 | S14 | S22 | S22 | S30 | S30 | S38 | S38 |
| E | Std 5 | Std 5 | S7 | S7 | S15 | S15 | S23 | S23 | S31 | S31 | S39 | S39 |
| F | Std 6 | Std 6 | S8 | S8 | S16 | S16 | S24 | S24 | S32 | S32 | S40 | S40 |
| G | S1 | S1 | S9 | S9 | S17 | S17 | S25 | S25 | S33 | S33 | S41 | S41 |
| H | S2 | S2 | S10 | S10 | S18 | S18 | S26 | S26 | S34 | S34 | S42 | S42 |

Example of plate layout for the Nitrite/Nitrate Assay Kit

7. Sample preparation

The following sample preparation protocols are intended as a guide only. The optimal conditions for sample preparation must be determined by the end user. It is recommended to use fresh samples. If it is not possible, aliquot and store samples appropriately with minimal freeze/thawing.

Nitrite/Nitrate Assay Kit can be used to determine nitrite in a wide variety of samples like biological fluids, cell lysates and tissue homogenates.

Biological fluids. Plasma and serum samples should be deproteinized by ultracentrifugation using a 10 kDa molecular weight cut-off filter. Citrate or EDTA are recommended as anticoagulants for plasma preparation. Urine can be used directly after dilution.

Tissue Homogenates. Dissect the tissue of interest and place it on a homogenizer tube with an appropriate amount of an ice-cold buffer. Homogenize the tissue and then centrifuge the homogenate at 10000 x g for 15 minutes at 4 °C and collect the supernatant.

Cell lysate. Wash cells with ice-cold buffer before lysis. Lyse cells by sonication or freeze-thaw cycles. Centrifuge cell lysis suspension at 10000 x g for 15 minutes at 4°C and collect the supernatant. It is recommended to use lysates from 1×10^6 cells.

Reagents and materials required for sample preparation are not supplied with the kit. Before doing sample preparation, consider the volume of sample required per test; see **Technical specifications** section.

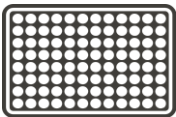







Make sure that interfering substances present in the sample do not give a significant background. Run proper blanks as necessary (e.g. sample blank should be always evaluated when working with highly colored samples). It is recommended to assay different sample dilutions to ensure the values fall within the standard curve.

8. Assay protocol

Prepare and mix all reagents thoroughly before use. Each standard, sample or blank should be assayed at least in duplicate.

It is possible to calculate both the **native nitrite** concentration and the **total nitrite** concentration (native nitrite + enzymatically reduced nitrate) in each sample

A. Total nitrite determination

-  Set up the plate design
-  Add **50 µL** of **standard** or **sample** in each well
-  Add **10 µL** of **R.A. Working Solution** and **20 µL** of **Reagent B** in each well
-  **Incubate** for **60 minutes** at RT
-  Add **10 µL** of **R.C. Working Solution** and **10 µL** of **Reagent D** in each well
-  **Incubate** for **20 minutes** at RT
-  Add **50 µL** of **Reagent E** in each well
-  **Incubate** for **10 minutes** protected from light at RT

9



Add **50 µL** of **Reagent F** in all wells

10



Incubate for **10 minutes** protected from light at **RT**

11



Read the **absorbance** of all wells at **540 nm** at **RT**

B. Native nitrite determination.

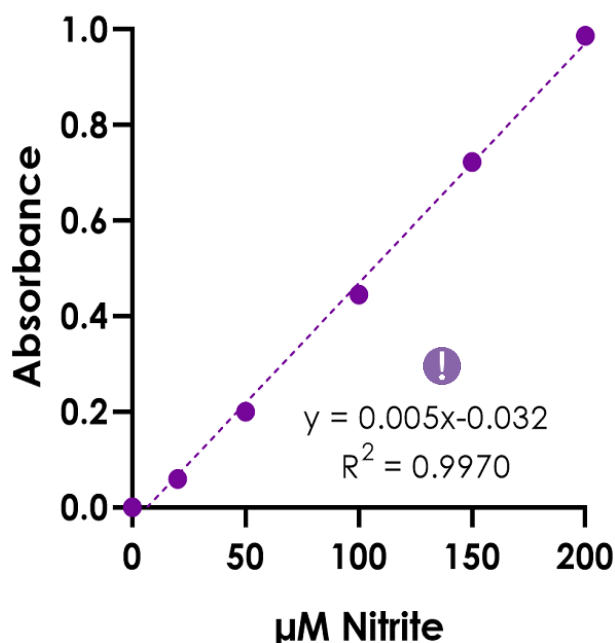
Follow the experimental procedure described for total nitrite determination in a sample **adding ddH₂O** to the wells instead of R.A. Working Solution, Reagent B, R.C. Working Solution and Reagent D (steps 3 and 5). Continue the assay by adding Reagents E and F as described in the protocol.

If you need to **adapt this kit** for another form of the assay (for example cuvette), **contact us at** info@bioquochem.com

9. Data analysis

ANALYSIS OF THE STANDARDS

- Calculate the average absorbance of the standards.
- Subtract the average absorbance of the reagent blank (Std 1) from the average absorbance of the standards to obtain the blank-corrected absorbance of the standards.
- Create a standard curve by plotting the blank-corrected absorbance of the standards as a function of the standard concentration (see **STANDARD CALIBRATION** section). A typical standard curve ($y = \text{slope} \cdot x \pm \text{intercept}$) for this assay is shown below.



Nitrite standard curve with Nitrite/Nitrate Assay Kit

- ⓘ This standard curve is an example of the data typically obtained with this kit. **DO NOT USE** this standard curve to calculate the nitrite content of your samples. A new standard curve must be performed by the end user.

ANALYSIS OF THE SAMPLES

- Calculate the average absorbance of the samples.
- Subtract the average absorbance of the reagent blank (Std 1) from the average absorbance of each sample to obtain the blank-corrected absorbance of the samples (A_s).
- Calculate the **nitrite concentration** of the samples using the following equation. Slope and intercept values are obtained from the standard curve.

$$\text{Total NO}_2^- (\mu\text{M}) = \left(\frac{A_s - \text{intercept}}{\text{slope}} \right)$$

When working with diluted samples the concentration values obtained must be multiplied by the dilution factor to obtain the nitrite content of the undiluted sample.

The results obtained depend on the assay format:

Total nitrite (nitrite+nitrate): concentration value obtained for the **total nitrite assay**.

Native nitrite: concentration value obtained for the **native nitrite assay**.

Nitrate: concentration obtained by **subtracting** the concentration calculated for **native nitrite from the total nitrite** concentration.

10. Troubleshooting

This troubleshooting table provides potential sources and solutions for common problems observed with BQC Assay Kits. **The problems listed below could occur when using any BQC Assay Kit.** They are not specific for this assay kit.

| Problem | Possible Cause | Recommended Solution |
|--|--|--|
| Wells have color but there is no reading | Plate read at incorrect wavelength | Check the wavelength used in the assay |
| | Incorrect microplate | Use the correct microplate for your application UV/Vis: transparent Fluorescence: black wells/transparent bottom |
| Standard readings do not follow a linear pattern | Pipetting errors in preparation of standards | Avoid pipetting small volumes (<5 μ L) Be careful not to splash from well to well |
| | Air bubbles formed in well(s) | Use reverse pipetting technique |
| | Standard stock is at incorrect concentration | Always refer to dilutions described in Assay preparation |
| | Improperly thawed reagents | Thaw all components completely and mix well before use |
| | Use of improperly stored reagents | Store the components appropriately Use fresh components from the standard curve |
| | Incorrect incubation times or temperatures | Refer to Assay protocol |
| Dispersion of standard and sample readings | Pipetting errors | Avoid pipetting small volumes (<5 μ L) Be careful not to splash from well to well |
| | Air bubbles formed in well(s) | Use reverse pipetting technique |

| Problem | Possible Cause | Recommended Solution |
|---|---|--|
| Sample erratic values | Samples contain interfering substances | Dilute sample further (if possible) |
| | Inappropriately stored samples or samples used after multiple freeze-thaw cycles | Use fresh samples or store appropriately until use |
| | Samples not deproteinized | Use an appropriate deproteinization protocol |
| | Cells/Tissue samples not homogenized completely | Repeat the sample homogenization |
| | Inappropriate sample dilution buffer | Refer to Assay preparation |
| Sample reading fall outside the detection range | Samples are too diluted/concentrated No analyte/activity is observed in the sample | Re-assay using different sample dilutions |

STILL HAVING PROBLEMS?

Contact BQC if you have any further questions, our team will be pleased to help you:



Phone

+ 34 985 26 92 92



E-mail

info@bioquochem.com


Business hours

 Monday-Thursday: 8.30 to 17.00 (CEST)
 Friday: 8.00 to 15.00 (CEST)

11. Additional information

Nitrite/Nitrate Assay Kit is a quick and precise assay (< 5 %) for nitrite determination in a wide variety of samples.

Antioxidants, azide, anticoagulants, dithiothreitol and mercaptoethanol have been reported to interfere with this assay.

If unexpected results are obtained running your samples, please contact us at info@bioquochem.com

12. Related products

More products available on bioquochem.com

| Reference | Product |
|-----------|-------------------------------------|
| KB03032 | Xanthine Oxidase Activity Assay Kit |
| KB03016 | MDA-TBARs Assay Kit |
| KB03009 | Nitrite Determination Assay Kit |

13. Warranties and limitation of liability

BQC shall not in any event be liable for incidental, consequential or special damages of any kind resulting from any use or failure of the products, even if BQC has been advised of the possibility of such damage including, without limitation, liability for loss of use, loss of work in progress, downtime, loss of revenue or profits, failure to realize savings, loss of products of buyer or other use or any liability of buyer to a third party on account of such loss, or for any labor or any other expense, damage or loss occasioned by such product including personal injury or property damage is caused by BQC's gross negligence. Any and all liability of BQC hereunder shall be limited to the amounts paid by the buyer for the product.

Buyer's exclusive remedy and BQC's sole liability hereunder shall be limited to a refund of the purchase price, or the replacement of all material that does not meet our specifications.

Said refund or replacement is conditioned on buyer giving written notice to BQC within 30 days of shipment.

Expiration date: 1 year from the date of fabrication. Expiration date is indicated on the outside of the box.

For further details, please refer to our website bioquochem.com



Edificio CEEI | Parque Tecnológico de Asturias,
33428 Llanera, Asturias
Info@bioquochem.com



www.bioquochem.com